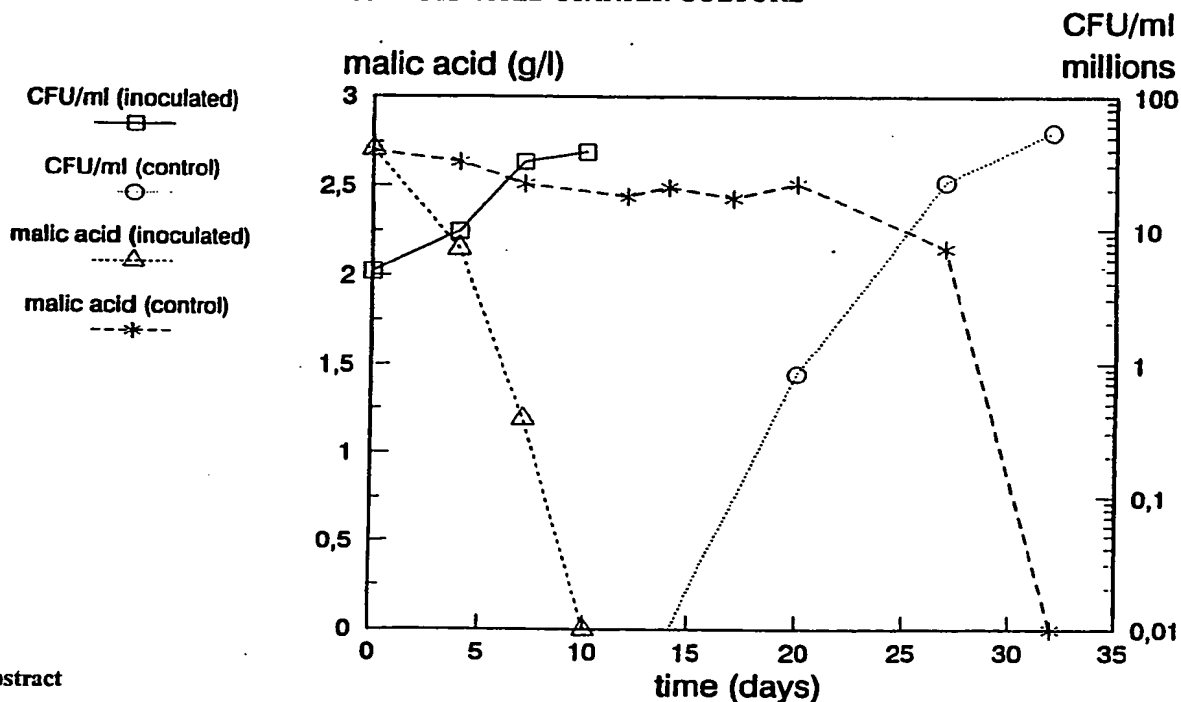




INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ : C12G 1/02, C12H 1/00 C12P 7/56		A1	(11) International Publication Number: WO 93/20180
			(43) International Publication Date: 14 October 1993 (14.10.93)
(21) International Application Number: PCT/DK93/00116		(81) Designated States: AU, NZ, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).	
(22) International Filing Date: 30 March 1993 (30.03.93)			
(30) Priority data: 07/864,823 1 April 1992 (01.04.92) US		Published <i>With international search report.</i>	
(71) Applicant (for all designated States except US): CHR. HANSEN'S LABORATORIUM DANMARK A/S [DK/DK]; Postboks 407, Bøge Allé 10-12, DK-2970 Hørsholm (DK).			
(72) Inventors; and (75) Inventors/Applicants (for US only) : PRAHL, Claus [DK/DK]; Mosevej 7, DK-3230 Græsted (DK). NIELSEN, Jan, Clair [DK/DK]; Lillebjergvej 4B, DK-3390 Hundested (DK).			
(74) Agent: PLOUGMANN & VINGTOFT A/S; Sankt Annæ Plads 11, Postboks 3007, DK-1201 Copenhagen K (DK).			

(54) Title: A METHOD OF INDUCING MALOLACTIC FERMENTATION IN WINE OR FRUIT JUICE BY DIRECT INOCULATION WITH A NON-ACTIVATED STARTER CULTURE



(57) Abstract

A method of inducing malolactic fermentation in wine or fruit juice by the direct inoculation of a concentrate of a starter culture containing a selected malolactically active bacterial strain having a survival rate of at least 80 % when introduced into a wine having a pH of 3.2 or lower and containing at least 25mg SO₂ per L and at least 12 vol % ethanol, and capable of starting malolactic fermentation when added directly to the wine or fruit juice at a concentration of less than 10⁷ colony forming units per mL.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	FR	France	MR	Mauritania
AU	Australia	GA	Gabon	MW	Malawi
BB	Barbados	GB	United Kingdom	NL	Netherlands
BE	Belgium	GN	Guinea	NO	Norway
BF	Burkina Faso	GR	Greece	NZ	New Zealand
BG	Bulgaria	HU	Hungary	PL	Poland
BJ	Benin	IE	Ireland	PT	Portugal
BR	Brazil	IT	Italy	RO	Romania
CA	Canada	JP	Japan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SK	Slovak Republic
CI	Côte d'Ivoire	LJ	Liechtenstein	SN	Senegal
CM	Cameroon	LK	Sri Lanka	SU	Soviet Union
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	MC	Monaco	TG	Togo
DE	Germany	MG	Madagascar	UA	Ukraine
DK	Denmark	ML	Mali	US	United States of America
ES	Spain	MN	Mongolia	VN	Viet Nam
FI	Finland				

A METHOD OF INDUCING MALOLACTIC FERMENTATION IN WINE OR FRUIT JUICE BY DIRECT INOCULATION WITH A NON-ACTIVATED STARTER CULTURE.

FIELD OF INVENTION

- 5 The present invention provides a novel method of inducing the decarboxylation of malic acid to lactic acid in wine or fruit juice by direct inoculation with a non-activated starter culture of malolactically active lactic acid bacteria and a composition comprising bacteria useful in the method.

10 BACKGROUND OF THE INVENTION

Grape juice, must and other fruit juice contain a varying amount of L-malic acid and L-malate, the amount typically being in the range of 1 to 10 g/L. The amount of malic acid and malate depends largely on the climatic conditions pre-
15 vailing in the viticultural region. Hence, wines produced in colder areas tend to have a relatively higher acid content, since the malic acid is not degraded during the normal alcoholic fermentation. From a taste and flavour point of view, malic acid is considered undesirable in most red wines and in
20 several types of rosé wines, white wines or sparkling wines.

However, the content of malic acid and malate in a wine may be reduced by a so-called malolactic fermentation (MLF) of the wine which fermentation results from the metabolic activity of various lactic acid bacteria, including species
25 belonging to the genera of Lactobacillus, Pediococcus and Leuconostoc. Such bacteria may be present in must and wine as part of the indigenous microbial flora hereof, or they may have been added as a bacterial starter culture. Typically, the MLF is associated with malolactic bacterial growth and
30 catabolic processes during which the wine acidity is reduced. The catabolic phase is usually entered when the malolactically active bacteria during the growth phase has reached a population density of about 10^6 colony forming units (CFU)

per mL. The microbial malolactic deacidification results from the decarboxylation of the dicarboxylic acid, L-malic acid to the monocarboxylic acid, L-lactic acid. As a result of this malolactic fermentation, the acidity of the wine decreases and the pH increases, resulting in a wine with a softer palate relative to that of the wine before the malolactic fermentation. Following a successful malolactic fermentation in wine, no further microbial growth will normally occur and hence, the wine is considered to be microbiologically stable.

10 The malolactic fermentation may occur spontaneously in the wine as a result of the growth of an indigenous flora of malolactically active lactic acid bacteria originating from the vines and grape skins and also often surviving from one season to the next on winery equipment, especially wooden
15 casks or other equipment made of wood. When occurring in this fashion, malolactic fermentation is often delayed and may take place several months after the alcoholic fermentation. The initial number of bacteria is often quite small and the environment of the wine is frequently rather hostile to the
20 growth of these bacteria due to the content of ethanol and sulphur dioxide in the wine, as well as its low pH and low nutrient concentration. The extended lag phase of the malolactic bacteria during which the wine is biologically unstable may result in the growth of bacteria producing volatile
25 acidity and hence spoilage of the wine. Apart from this, certain indigenous malolactic bacteria spontaneously growing in the wine may produce certain compounds, e.g. biogenic amines that are believed to give rise to health problems.

In the traditional winery industry where the spontaneously occurring malolactic fermentation is being relied upon, it is
30 common practice to stimulate malolactic fermentation by reducing the amount of added sulphur when determined as sulphur dioxide, to below 50 mg per L, delaying the removal of the lees, increasing the temperature of the wine to above
35 20°C, or ensuring a pH of more than 3.4. These measures,

however, may also favour the growth of undesired microorganisms in the wine such as Acetobacter species, thus increasing the likelihood of wine spoilage, and this approach therefore requires extremely careful supervision of the

5 decarboxylation process. Even if precautions are taken to enhance spontaneous MLF, this process is still difficult, not to say impossible, to control, and its occurrence has become even more unpredictable as winery hygiene has improved e.g. as a result of the replacement of wooden casks with stainless

10 steel tanks. Such hygienic measures serve to ensure a uniform quality of the wine and reduce the risk of spoilage. However, they also reduce the chance of spontaneous conversion of malic acid taking place in the wine.

For this reason, and because the winemaker will often prefer

15 to exercise a greater degree of control over the malolactic fermentation process it has become increasingly common practice in the winery industry to add a starter culture of malolactically active bacteria to the wine after the alcoholic fermentation. Presently, such a postfermentation malolac-

20 tic fermentation in wine may be induced in different ways.

Currently, a widely practiced method of inducing MLF is to seed a wine with a small proportion of another wine already undergoing MLF and thus containing a high number of viable malolactically active bacteria. The culture in the seed wine

25 is then already well adapted to wine conditions and will usually be capable of completing the malolactic fermentation in the seeded wine. However, this method of inoculation is rather tedious and not completely controllable. Thus, the method requires that a concentrated "mother culture" of the

30 bacteria is propagated for an extended period of time, such as about two months, in diluted wine or grape juice, optionally after rehydration and/or adaptation of the mother culture in e.g. a grape fruit juice-containing medium which is then used to inoculate the wine to be used as a bulk

35 starter culture in the form of seed wine. Typically, the wine is inoculated with the seed wine at a rate of 1 to 10 vol%,

and accordingly, this method requires substantial investments in propagation equipment and adequately trained staff. In addition, it is difficult to control the propagation process and hence serious timing problems may occur. Should both red
5 and white wine starter cultures be required these problems are doubled.

Relative to the above practice of using a seed wine, a post-fermentation inoculation of wine with concentrated freeze-dried or frozen starter cultures of malolactically active
10 lactic acid bacteria implies considerable savings in time and labour by substantially reducing the amount of scale-up work required. Such concentrated starter cultures are now commercially available and their use is being increasingly accepted in the wine industry, although their malolactic effect is not
15 completely reliable. These culture compositions may be concentrated to contain a number of colony forming units which, when determined in a non-inhibitory medium is in the range of 10^9 to 10^{11} per g of culture.

However, the use of known, commercially available malolactically active culture compositions contain bacteria which
20 are susceptible to the conditions (low pH, content of SO_2 , content of ethanol) prevailing in the fermented wine and accordingly, their malolactic efficiency requires that they become adapted to the hostile conditions in the wine by a
25 carrying out a thawing and rehydration step (if freeze-dried), a thawing step (if frozen) and an activation step prior to inoculation in the wine. This requirement is assumingly due to the well-known phenomenon that a freeze-dried or frozen lactic acid bacterial culture composition, even if
30 it includes additives protecting the bacteria against cell damages, will as a result of the freezing and/or freeze-drying process have an increased susceptibility to low pH, SO_2 , ethanol and low temperatures as compared to a freshly grown culture of the same species.

If not rehydrated and activated as described above, the survival rate of the known commercial malolactic compositions on direct inoculation into wine will typically be in the range of 0.01 to 1% or even lower. Furthermore, the initially surviving non-adapted bacteria may gradually lose their viability in the wine.

Typically, this required adaptation comprises an initial thawing and/or rehydration process, the latter comprising dissolving the freeze-dried composition in water and adding various nutrients such as a sugar, vitamins, minerals or yeast extract and keeping the resulting solution at about 22°C for about one hour. Subsequently, the thus rehydrated composition is subjected to an activation step, typically lasting 48 to 84 hours, in a medium which typically comprises grape juice or wine diluted with water, yeast extract, trace elements and vitamins. Normally, the number of CFUs does not increase during this activation period, on the contrary, the number may decrease. Even if the starter culture composition is adapted as described above, a varying proportion of adapted bacteria may lose viability as determined in a non-inhibitory medium when inoculated in wine. This loss of viability may be up to 90% of the added number of CFUs.

In EP-A1-0141878 is disclosed a method of reducing malic acid to lactic acid in wine by the introduction of high numbers of bacterial cells into the wine, which method comprises activating a concentrate of a bacterial culture in a nitrogen source-augmented fruit juice to form an activated mixture of bacteria containing at least about 10^5 CFUs per mL and introducing the activated mixture into wine or grape must and converting the malic acid to lactic acid. The activation conditions disclosed are an activation period of 48 hours at 24°C.

Krieger et al. (Food Biotechnol. 1990, 7, 484) have disclosed the application in wine of malolactically active strains of Leuconostoc oenos and Lactobacillus spp. in the form of fresh

or frozen concentrates using direct inoculation, i.e. without preceding activation, of at least 10^7 CFUs per mL which concentration is indicated as being necessary to start malolactic fermentation. However, the application of such a high concentration of malolactically active organisms is not commercially feasible in the wine industry due to the high cost of starter cultures. The necessity to apply at least 10^7 CFUs per mL as disclosed in order to start MLF may indicate that the survival rate of these organisms when applied directly to the wine is so low that a malolactically active concentration of the organisms is only achieved at the indicated inoculation level.

The present invention provides, compared to the known methods, a significantly improved method of inducing malolactic fermentation in wine or fruit juice whereby it has become possible to achieve an effective malolactic fermentation herein within a short period of time by inoculating wine or a fruit juice directly with a concentrated culture composition of malolactically active bacteria at an economically feasible concentration and accordingly, to avoid the tedious and costly processes of rehydration, activation, adaptation and/or expansion which are currently required with commercial malolactically active starter cultures.

SUMMARY OF THE INVENTION

Accordingly, the present invention relates in a first aspect to a method of inducing in a wine or a fruit juice the conversion of malic acid to lactic acid comprising the steps of selecting a strain of malolactically active bacteria, capable of starting malolactic fermentation in a wine or fruit juice when added directly thereto at a concentration of less than 10^7 CFUs per mL, providing a concentrate of the selected malolactically active bacterium, introducing said concentrate directly into the wine or the fruit juice, and converting the malic acid while keeping the wine or the fruit juice under

conditions which allow conversion of the malic acid to obtain a malolactically fermented wine or fruit juice having a content of malic acid which is at the most 0.5 g per L.

In a further aspect, the invention provides a selected strain
5 of a malolactically active bacterium which, when it is added to a wine or fruit juice in a frozen or freeze-dried state at a concentration of CFUs which is in the range of 1×10^6 to 5×10^7 per mL of the wine or fruit juice, is capable of reducing at least 4 g of malic acid per L of wine or fruit juice
10 to less than 0.5 g per L within a period of time which is at the most 15 days, said strain having at least one of following characteristics:

(a) capable of decarboxylating malic acid in wine at a pH of 3.2 or lower,

15 (b) capable of decarboxylating malic acid in wine in the presence of 25 mg SO₂ per L or more, and

(c) capable of decarboxylating malic acid in wine having an ethanol content of 10 vol% or higher.

20 (d) a survival rate of at least 80% when introduced directly into a wine having a pH of 3.2 or lower and containing at least 10 vol% ethanol,

(e) a survival rate of at least 50% when introduced directly into a wine having a pH of 3.2 or lower and containing at least 25 mg SO₂ per L and at least 12
25 vol% ethanol,

(f) resistant to attack by bacteriophages,

(g) capable of retaining at least one of the characteristics (a) through (f) during propagation and concentration.

In a still further aspect, the invention relates to a concentrate of one or more of the selected strains of malolactically active bacteria as defined herein, and in another aspect there is provided a malolactic starter culture composition comprising a concentrate of selected malolactically active bacterial strain(s), as defined herein and at least one further ingredient selected from cryoprotectants, bacterial nutrients and bulking agents.

DETAILED DISCLOSURE OF THE INVENTION

- 10 The present invention relates in one aspect to a novel method of inducing the decarboxylation of malic acid to lactic acid in wines and fruit juices by direct inoculation of the wine or fruit juice with appropriately selected malolactically active bacteria.
- 15 In the present context, the term "wine" is used to describe a product resulting from an alcoholic fermentation of juice or must of grapes or of any other fruit or berries, whether the fermentation occurs spontaneously or it is obtained by the addition of a yeast culture.
- 20 A wine made from grapes may be a red wine, a white wine, a rosé wine, all of which may be in the form of sparkling wines.

The conditions prevailing in wine at the completion of the alcoholic fermentation is generally unfavorable for microbial growth. These adverse conditions include a pH typically being in the range of 2.8 to 4.0 such a pH of 3.2 or lower, an ethanol content which is typically in the range of 8 to 14 vol%, such as in the range of 10 to 12 vol%, a low content of nutrients such as carbon sources due to the depletion of the nutrients during the alcoholic fermentation. In addition, a sulphur-containing substance may have been added as a preservative to the must in an amount which is typically in the

range of 5 to 70 mg per L including amounts of 25 mg of SO₂ as defined below per L in the fermented wine, or higher.

In this connection it should be noted that the addition of sulphur-containing substances to must is conventionally carried out by the addition of a sulphite or another water soluble sulphur-containing substance. In the fermenting must the sulphite is at least partially converted to SO₂, the extent of the conversion depending i.a. on the pH. Part of the generated SO₂ will be in the form of free molecules and part will be bound. In the art, the content of sulphur in must or wine is determined by measuring the total content of sulphur-containing substances as total SO₂. Accordingly, when used herein the term "SO₂" denotes the total content of sulphur determined according to the Rebelein method in which all sulphur-containing substances are converted to SO₂ before the measuring step.

These conditions in the wine, singly and in combination provides a rather hostile environment for bacteria, including malolactically active lactic acid bacteria. Such bacteria are primarily organisms belonging to the genera of Leuconostoc, Lactobacillus and Pediococcus. Among these genera of naturally occurring malolactically active bacteria, organisms belonging to Leuconostoc are generally most tolerant to low pH and they may grow at pH values below 3.3. At more moderate pH values such as pH of 3.6 and above, species of Lactobacillus and Pediococcus may grow as well. Within the Leuconostoc genus, the species Leuconostoc oenos is particularly adapted to grow in wines and this species is commonly used in commercial cultures as described above. Species of Lactobacillus which are of particular interest as malolactic starter culture organisms include Lactobacillus casei, Lactobacillus brevis, Lactobacillus hilgardii and Lactobacillus plantarum.

However, within a species of malolactically active bacteria there may exist differences between different isolates as to

tolerance to one or more of the above-mentioned adverse conditions in wine. Accordingly, it may be possible to select, within a species of malolactically active bacteria, isolates (strains) which are particularly tolerant to one or more of the adverse conditions. However, it is known in the art that when selecting a bacterial strain against one desired characteristic, such a strain will frequently be industrially less useful due to deficiencies as to other desired characteristics. E.g. a strain selected on the basis of a high tolerance to acidic conditions be susceptible to moderate amounts of ethanol.

In a first step of the presently claimed method, a selection is carried out to obtain a strain of malolactically active bacteria, capable of starting malolactic fermentation in wine or fruit juice when added directly thereto at a concentration of less than 10^7 CFUs per mL, the strain having at least one of the following characteristics:

- (a) capable of decarboxylating malic acid in wine at a pH of 3.2 or lower,
- (b) capable of decarboxylating malic acid in wine in the presence of 25 mg SO_2 per L or more,
- (c) capable of decarboxylating malic acid in wine having an ethanol content of 10 vol% or higher,
- (d) a survival rate of at least 80% when introduced into a wine having a pH of 3.2 or lower and containing at least 10 vol% ethanol,
- (e) a survival rate of at least 50% when introduced into a wine having a pH of 3.2 or lower and containing at least 25 mg SO_2 per L and at least 12 vol% ethanol,
- (f) resistant to attack by bacteriophages, or

(g) capable of retaining at least one of the characteristics (a) through (f) during propagation and concentration.

When the selection criteria as those defined above have been
5 determined, appropriate selection procedures known per se in
the art can readily be chosen. Initially, a large number of
strains of malolactically active bacteria are isolated from
different types of wines and preferably from a range of
viticultural regions. Such isolates may subsequently be
10 tested individually according to each of the above selection
criteria, or a number of the isolates may more conveniently
be combined and subjected to a first selection in a wine used
as a growth medium in a turbidostat fermentor to obtain
selection of strains which are capable of growing in a wine
15 at a low pH and a high ethanol content.

When using such a turbidostat growth selection procedure, the
mixture of isolates are inoculated into a fermentor and the
biomass is kept constant by means of a turbidometer. The
culture is fed a nutrient-enriched wine with an increasing
20 concentration of ethanol and a decreasing pH. When the pH
level and the ethanol concentration is so high that no fur-
ther bacterial growth is possible, a sample of the culture is
drawn and representative strains of viable malolactically
active bacteria are isolated and characterized.

25 In a second selection step, the isolated, acid and ethanol
tolerant strains are tested in wines having different compo-
sitions for the characteristics (a) through (f) as mentioned
above. In a next step, chosen strains are subjected to fer-
mentor propagation under normal production conditions in
30 order to select strains which have an industrially feasible
growth yield and which at the same time maintain the charac-
teristics based on which they were selected.

In a final selection step, the strains which pass the prece-
ding step is subjected to a down-stream process including at

least one of the following steps: harvesting of cells from the culture medium, concentration of the cells, freezing of the concentrate or freeze-drying.

It will be understood that a selected strain should preferably have as many as possible of the above-defined characteristics (a) through (g). Accordingly, the selected strain is preferably one which has at least two of the characteristics, more preferably one having at least three of the characteristics and most preferably the strain is one which has all of the defined characteristics. In particular, the selected strain should in addition to other advantageous characteristics have a high rate of survival under the conditions as defined above. A high survival rate makes it possible to have the MLF started at the low CFU concentrations as defined above and allow the high proportion of surviving cells to enter the active growth phase and becoming malolactically active essentially momentarily or within a short period of time such as within 1-3 days, thereby obtaining the malolactically fermented wine or fruit juice within the short periods of time as defined herein. It may be observed with cultures having a low survival rate and applied in high concentrations that although an initial conversion of malic acid takes place due to the release of malolactically active enzymes from the killed cells, the active growth phase is entered only after a prolonged lag phase of 5-10 days and accordingly, such cultures will not be suitable for obtaining a rapid MLF and particularly, these cultures will not result in a conversion of high amounts of malic acid within an industrially acceptable period of time, such as it is obtained with the present selected strains.

In accordance with the invention, a suitable malolactically active bacterial strain is one selected from a species belonging to the genera of Leuconostoc, Lactobacillus and Pediococcus. When the strain is selected from a species of the Leuconostoc genus, the species is preferably Leuconostoc oenos such as a strain which is selected from the group

consisting of DSM 7008, DSM 7009, DSM 7010, DSM 7011, DSM 7012, DSM 7013, DSM 7014 and DSM 7015. These 8 strains were deposited on 26 March, 1992 with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (German Collection of
5 Microorganisms and Cell Cultures), Braunschweig, Germany.

In a second step of the method as claimed herein, there is provided a concentrate of the thus selected malolactically active strain. The concentrate is preferably one in which the number of colony forming units is in the range of 10^9 to 10^{12}
10 per g. In more preferred embodiments, the concentrate has a number of colony forming units which is in the range of 10^{11} to 10^{12} per g.

As an initial step in the obtainment of the concentrate, a chosen selected strain is propagated according to processes
15 which are well-known in the art. Such processes include the step of cultivating the strain in a suitable fermentor vessel containing a suitable growth medium containing sufficient amounts of nutrients required for economically feasible growth yield of the particular strain. In accordance with the
20 invention, a suitable growth medium for propagation of malolactically active bacteria contains as the major ingredient grape juice to which may be added yeast concentrate or extract, malic acid, a surface active substance such as Tween™ 80, a manganese salt, water and a vitamin mixture. In preferred
25 embodiments of the invention, the pH in the growth medium is in the range of 4 to 6.

Subsequent to the above propagation step, the cell biomass is harvested e.g. by centrifugation whereby a concentrate of cells is obtained in the form of a slurry or a cell paste.
30 Alternatively, the concentrate of cells may be obtained by a filtration process. Although such a fresh cell paste may be used to inoculate a wine or fruit juice, the bacterial concentrate is more conveniently provided in the form of a preserved concentrate, since the viability of cells in a cell
35 paste concentrate will decrease rapidly and furthermore, a

fresh cell paste will be prone to growth of contaminating microorganisms. In accordance with the invention, a concentrate may be preserved by freezing e.g. by dripping the paste into liquid nitrogen or by introducing the cell paste into a suitable freezing apparatus. In order to retain the viability of cells during freezing, the paste may be mixed with suitable cryoprotectants such as gelatine, prior to freezing. In a preferred embodiment of the invention, the concentrate is provided as a freeze-dried concentrate which is typically prepared by subjecting a frozen concentrate to a conventional freeze-drying process.

In a subsequent step of the claimed method, a concentrate of the selected malolactically active strain is inoculated directly into the malic acid-containing wine or fruit juice. In the present context, the term "inoculated directly" is used to describe that the concentrate as defined herein is added to the wine or fruit juice without any prior rehydration and/or activation step. As it has been explained above, the known commercial malolactic starter cultures for post-alcoholic MLF all require that they are subjected to an adaptation step including an activation step and optionally (if they are in a freeze-dried state) an initial rehydration step, prior to inoculation. If such known cultures are inoculated directly, the viability of the cells is typically reduced by a factor of 100 to 1000.

The concentrate is added to the wine or the fruit juice in an amount which results in the introduction of a number of bacteria, calculated on a CFU basis which is in the range of 10^5 to 10^7 per mL. The preferred inoculation rate depends on several factors including the amount of malic acid to be converted, the desired period for obtaining the malolactically fermented wine or fruit juice and the temperature conditions. A typical preferred inoculation rate will be in the range of 10^6 to 10^7 CFUs per mL and a more preferred range may be 5×10^6 to 1×10^7 CFUs per mL.

In the final step of the method as claimed herein, the malic acid is converted to lactic acid while keeping the inoculated wine or fruit juice under conditions which will allow the conversion to take place. In particular, it is required to
5 keep the wine or the fruit juice at a temperature where the inoculated culture is malolactically active. The majority of malolactic bacteria will be active at temperatures in the range of about 12 to about 25°C. A typical temperature for storing wine undergoing malolactic fermentation will be in
10 the range of about 15 to 22°C.

As mentioned above, the content of malic acid in a wine may vary, in particular according to the climatic conditions of the viticultural region. Typically, the malic acid content will be in the range of 2 to 10 g per L. In specific embo-
15 diments of the invention, the wine or fruit juice to be malolactically fermented as defined herein is one having a malic acid content of at least 4 g per L, such as at least 5.5 g per L. In accordance with the invention, a malolactically fermented wine or fruit juice containing less than 0.5
20 g malic acid per L should preferably be obtained from such a wine or fruit juice within a period of time which is at the most 15 days. In more preferred embodiments, the period of time is at the most 12 days and in still more preferred embodiments it is at the most 10 days such as at the most 8
25 days.

In particularly preferred embodiments of the present invention, the obtained malolactically fermented wine or fruit juice has a malic acid content which is at the most 0.1 g per L.

30 As mentioned above, the present invention relates in further aspects to a strain of a malolactically active bacterium which has been selected according to the methods and selection criteria as defined above and to a concentrate of such a selected strain as also defined above.

Finally, the present invention provides a malolactic starter culture composition comprising the concentrate as defined herein and at least one further ingredient selected from cryoprotectants, including as an example gelatine, bacterial
5 nutrients and bulking agents.

Conveniently, the claimed composition is preserved to obtain a commercial product which has a suitable shelf-life when stored and distributed appropriately. Accordingly, the composition is suitably a frozen or a freeze-dried composition
10 which has been prepared by subjecting a mixture of the bacterial concentrate and the further ingredient(s) to a freezing and/or freeze-drying process essentially in accordance with the above-mentioned processes used for the claimed concentrate.

15 Preferably, the composition is one having a content of colony forming units which is in the range of 10^9 to 10^{12} per g, such as in the range of 10^{11} to 10^{12} per g.

In useful embodiments of the invention, the composition is one which when it is inoculated directly into a wine or a
20 fruit juice containing at least 4 g of malic acid per L such as at least 5.5 g per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time which is at the most
25 15 days, preferably at the most 12 days and more preferably at the most 10 days such as at the most 8 days.

It may be advantageous to provide the claimed composition as a composition comprising a multiplicity of the hereindefined selected strains of malolactically active bacteria. Accordingly, in one embodiment the composition comprises such a
30 multiplicity.

DESCRIPTION OF DRAWINGS

The invention is further explained below with reference to the drawings in which

Fig. 1 shows the survival and growth (%CFU/mL) of Leuconostoc
5 oenos strain DSM 7008 as fresh culture (fermentate) and as processed concentrates after direct inoculation into wine made from Riesling grape juice (11.5 vol% ethanol, 0 mg SO₂ per L, pH 3.15) of 3 x 10⁶ to 7 x 10⁶ per mL.

Fig. 2A shows the conversion of malic acid and CFU/mL of
10 malolactically active bacteria in Chénin white wine (10.5 vol% ethanol, 16 mg SO₂/mL, pH 3.5, 0.004 g/L of glucose and 0.003 g/L of fructose) kept at 18°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc
15 oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2B shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Sauvignon red wine (11.0 vol% ethanol, 0 mg SO₂/mL, pH 3.6, 0.11 g/L of glucose and 0.07 g/L of fructose) kept at 20°C in 5 L jars inoculated
20 with a freeze-dried composition of Leuconostoc oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2C shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Sauvignon red wine (11.3 vol% ethanol, 0 mg SO₂/mL, pH 3.6, 0.11 g/L of glucose
25 and 0.07 g/L of fructose) kept at 21°C in 10.000 L insulated tanks inoculated with a freeze-dried composition of Leuconostoc oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2D shows the conversion of malic acid and CFU/mL of
30 malolactically active bacteria in Cabernet Sauvignon red wine (11.8 vol% ethanol, 5 mg SO₂/mL, pH 3.5, 0.3 g/L of glucose and 0.45 g/L of fructose) kept at 20°C in 5 L jars inoculated

with a freeze-dried composition of Leuconostoc oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2E shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Sauvignon red wine (11.8 vol% ethanol, 5 mg SO₂/mL, pH 3.5, 0.3 g/L of glucose and 0.45 g/L of fructose) kept at 20°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2F shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Sauvignon red wine (11.0 vol% ethanol, 0 mg SO₂/mL, pH 3.6, 0.11 g/L of glucose and 0.07 g/L of fructose) kept at 20°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc oenos strain DSM 7015, and in the same wine without inoculation.

Fig. 2G shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Sauvignon red wine (11.8 vol% ethanol, 5 mg SO₂/mL, pH 3.5, 0.3 g/L of glucose and 0.45 g/L of fructose) kept at 20°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc oenos strain DSM 7015, and in the same wine without inoculation.

Fig. 2H shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Cabernet Franc red wine (13.0 vol% ethanol, 2 mg SO₂/mL, pH 3.4) kept at 20°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc oenos strain DSM 7008, and in the same wine without inoculation.

Fig. 2I shows the conversion of malic acid and CFU/mL of malolactically active bacteria in Semillon white wine (12.2 vol% ethanol, 26 mg SO₂/mL, pH 3.2) kept at 19°C in 5 L jars inoculated with a freeze-dried composition of Leuconostoc oenos strain, DSM 7008, and in the same wine without inoculation.

EXAMPLE 1

Survival and growth of Leuconostoc oenos in an experimental wine

An experimental wine was prepared by yeasting a sterile
5 Riesling grape juice originating from Germany, without the
addition of sulphite. After completion of the alcoholic
fermentation, the wine was sterile filtered and the resulting
wine had the following composition:

	Ethanol	11.5 vol%
10	Malic acid	3.9 g/L
	Residual sugar	3.5 g/L
	pH	3.15

A culture of Leuconostoc oenos strain DSM 7008 was prepared,
and the sterile wine was inoculated with (1) cells of the
15 strain directly from the fermentate (i.e. the growth medium
containing an outgrown culture of the strain), (2) a cell
concentrate after centrifugation, (3) a cell concentrate to
which cryoprotective agents had been added or (4) a freeze-
dried composition of a cell concentrate with added cryopro-
20 tectants, respectively. The numbers of CFUs which were inocu-
lated by the four mentioned forms of the strain varied
between 3×10^6 and 7×10^6 per mL of the experimental wine.

The survival of the added bacteria in the wine kept at 20°C
was monitored over 3 days, samples being collected on day 2
25 and 3 and the number of CFUs herein being determined accor-
ding to standard methods for determining viable counts of
Leuconostoc oenos.

The results of this experiment are shown in Fig. 1 from which
it appears that the bacteria when added in the form of a
30 fermentate multiplied during the keeping time, with a factor
of about 6. Bacteria added in the form of processed inocula
showed a survival rate after 2 days which was in the range of

80% to 100% and during the next day the numbers of CFUs of these inocula increased by 10% to 60% relative to the initial numbers.

EXAMPLE 2

5 Induction of malolactic fermentation in Bordeaux red wines and white wines by direct inoculation with *Leuconostoc oenos*-containing compositions

10 The malolactic effect of freeze-dried compositions of two strains of *Leuconostoc oenos*, DSM 7008 and DSM 7015 were tested by direct inoculation into different wines. The study was carried out by inoculating wines in 5 L jars and in industrial scale by inoculating wines contained in 5.000 to 10.000 L tanks. The inoculum level was about 5×10^6 per mL of wine.

15 The study comprised the following experiments:

1. Inoculation with strain DSM 7008 of Chénin white wine from the Loire region contained in 5 L jars.
2. Inoculation with strains DSM 7008 and DSM 7015 of Cabernet Sauvignon red wine from Graves in the Bordeaux region, kept in 5 L jars and additionally with DSM 7008 in a 10.000 L tank.
- 25 3. Inoculation with strains DSM 7008 and DSM 7015 of Cabernet Sauvignon red wine from Premières Côtes de Bordeaux contained in 5 L jars and additionally with strain DSM 7008 in a 5.000 L tank. This wine had a content of ethanol of 11.8 vol% and a SO₂ content of 5 mg per L and a pH of 3.5.
4. Inoculation with strain DSM 7008 of Cabernet-Franc red wine from Premières Côtes de Bordeaux, kept in 5 L jars.

5. Inoculation with strain DSM 7008 of Semillon white wine from Côtes de Montravel in the Bordeaux region, kept in 5 L jars.

The tested wines had an initial content of malic acid in the range of 3 to 5.5 g per L, and the content of SO₂ was in the range of 0 to 26 mg per L. In all of the inoculated wines the malolactic fermentation occurred at a rate which resulted in a substantially complete disappearance of malic acid within 8 to 20 days. The survival rate of the inoculated bacteria was 90 to 100% in the test wines and in most wines an increase of CFUs up till about 10⁸ per mL occurred during the malolactic fermentation period.

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT <div style="text-align: right;">Further deposits are identified on an additional sheet <input checked="" type="checkbox"/></div>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7011
C. ADDITIONAL INDICATIONS (leave blank if not applicable) <div style="text-align: right;">This information is continued on an additional sheet <input checked="" type="checkbox"/></div>	
<p>As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.</p>	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

<div style="text-align: center;">For receiving Office use only</div> <div style="border: 1px solid black; padding: 5px;"> <input checked="" type="checkbox"/> This sheet was received with the international application </div> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> </div> <div style="border: 1px solid black; padding: 5px; margin-top: 5px;"> Authorized officer </div>	<div style="text-align: center;">For International Bureau use only</div> <div style="border: 1px solid black; padding: 5px;"> <input type="checkbox"/> This sheet was received by the International Bureau on: </div> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;"> </div> <div style="border: 1px solid black; padding: 5px; margin-top: 5px;"> Authorized officer </div>
---	---

Form PCT/RO/134 (July 1992)

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>2</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution <div style="margin-left: 40px;">DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH</div>	
Address of depositary institution (including postal code and country) <div style="margin-left: 40px;">Mascheroder Weg 1 B D-3300 Braunschweig</div>	
Date of deposit 26 March 1992	Accession Number DSM 7012
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="text-align: center;">For receiving Office use only</div> <div style="border: 1px solid black; padding: 5px;"><input checked="" type="checkbox"/> This sheet was received with the international application <div style="margin-left: 100px;"><i>Anne Ryding</i></div><div style="border-top: 1px solid black; margin-top: 5px;">Authorized officer</div></div>	<div style="text-align: center;">For International Bureau use only</div> <div style="border: 1px solid black; padding: 5px;"><input type="checkbox"/> This sheet was received by the International Bureau on: <div style="border-top: 1px solid black; margin-top: 5px;">Authorized officer</div></div>

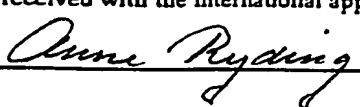
INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>2</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7013
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="text-align: right; margin-bottom: 5px;">For receiving Office use only</div> <div style="border: 1px solid black; padding: 5px;"><input checked="" type="checkbox"/> This sheet was received with the international application <div style="text-align: center; margin-top: 10px;"></div><div style="border-top: 1px solid black; margin-top: 5px;">Authorized officer</div></div>	<div style="text-align: right; margin-bottom: 5px;">For International Bureau use only</div> <div style="border: 1px solid black; padding: 5px;"><input type="checkbox"/> This sheet was received by the International Bureau on: <div style="border-top: 1px solid black; margin-top: 10px;">Authorized officer</div></div>

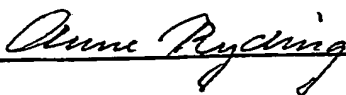
INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>2</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7014
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="text-align: center; border-bottom: 1px solid black; margin-bottom: 5px;">For receiving Office use only</div> <div style="border: 1px solid black; padding: 5px;"><input checked="" type="checkbox"/> This sheet was received with the international application <div style="text-align: center; margin-top: 10px;"></div><div style="border-top: 1px solid black; margin-top: 5px;">Authorized officer</div></div>	<div style="text-align: center; border-bottom: 1px solid black; margin-bottom: 5px;">For International Bureau use only</div> <div style="border: 1px solid black; padding: 5px;"><input type="checkbox"/> This sheet was received by the International Bureau on: <div style="border-top: 1px solid black; margin-top: 10px;">Authorized officer</div></div>

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

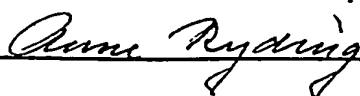
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>2</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7015
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="border: 1px solid black; padding: 5px;"><div style="text-align: center; font-weight: bold; font-size: small;">For receiving Office use only</div><div style="display: flex; justify-content: space-between; align-items: center;"><input checked="" type="checkbox"/> This sheet was received with the international application</div><div style="text-align: center; margin-top: 10px;"> Authorized officer</div></div>	<div style="border: 1px solid black; padding: 5px;"><div style="text-align: center; font-weight: bold; font-size: small;">For International Bureau use only</div><div style="display: flex; justify-content: space-between; align-items: center;"><input type="checkbox"/> This sheet was received by the International Bureau on:</div><div style="text-align: center; margin-top: 10px;">Authorized officer</div></div>

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

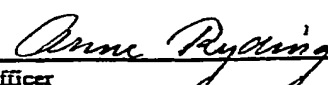
(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7008
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

<div style="text-align: right; font-weight: bold; font-size: small;">For receiving Office use only</div> <div style="padding: 5px;"><input checked="" type="checkbox"/> This sheet was received with the international application</div> <div style="text-align: center; margin-top: 20px;"></div> <div style="border-top: 1px solid black; padding-top: 5px;">Authorized officer</div>	<div style="text-align: right; font-weight: bold; font-size: small;">For International Bureau use only</div> <div style="padding: 5px;"><input type="checkbox"/> This sheet was received by the International Bureau on:</div> <div style="border-top: 1px solid black; padding-top: 5px;">Authorized officer</div>
--	---

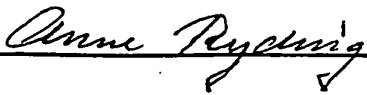
INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7009
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="border: 1px solid black; padding: 5px;"><div style="text-align: center; font-weight: bold; font-size: small;">For receiving Office use only</div><div style="display: flex; justify-content: space-between; align-items: center;"><input checked="" type="checkbox"/> This sheet was received with the international application</div><div style="text-align: center; margin-top: 20px;"> Authorized officer</div></div>	<div style="border: 1px solid black; padding: 5px;"><div style="text-align: center; font-weight: bold; font-size: small;">For International Bureau use only</div><div style="display: flex; justify-content: space-between; align-items: center;"><input type="checkbox"/> This sheet was received by the International Bureau on:</div><div style="text-align: center; margin-top: 20px;">Authorized officer</div></div>

INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>13</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSM Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1 B D-3300 Braunschweig	
Date of deposit 26 March 1992	Accession Number DSM 7010
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input checked="" type="checkbox"/>	
As regards the respective Patent Offices of the respective designated states, the applicants request that a sample of the deposited microorganisms only be made available to an expert nominated by the requester until the date on which the patent is granted or the date on which the application has been refused or withdrawn or is deemed to be withdrawn.	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	
<div style="text-align: right; margin-bottom: 5px;">For receiving Office use only</div> <div style="border: 1px solid black; padding: 5px;"><input checked="" type="checkbox"/> This sheet was received with the international application</div> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;">Authorized officer <div style="text-align: center; margin-top: 10px;"></div></div>	<div style="text-align: right; margin-bottom: 5px;">For International Bureau use only</div> <div style="border: 1px solid black; padding: 5px;"><input type="checkbox"/> This sheet was received by the International Bureau on:</div> <div style="border: 1px solid black; padding: 5px; margin-top: 10px;">Authorized officer</div>

CLAIMS

1. A method of inducing in a wine or a fruit juice the conversion of malic acid to lactic acid comprising the steps of selecting a strain of malolactically active bacteria, capable
5 of starting malolactic fermentation in a wine or fruit juice when added directly thereto at a concentration of less than 10^7 CFUs per mL, providing a concentrate of the selected malolactically active bacterium, introducing said concentrate directly into the wine or the fruit juice, and converting the
10 malic acid while keeping the wine or the fruit juice under conditions which allow conversion of the malic acid, to obtain a malolactically fermented wine or fruit juice having a content of malic acid which is at the most 0.5 g per L.
2. A method according to claim 1 wherein the wine or the
15 fruit juice has a pH which is at the most 3.2.
3. A method according to claim 1 wherein the wine is a red wine.
4. A method according to claim 1 wherein the wine is a wine selected from a white wine and a sparkling wine.
- 20 5. A method according to claim 1 wherein the malolactically active bacterial strain is one selected from the group consisting of a Leuconostoc species, a Lactobacillus species and a Pediococcus species.
6. A method according to claim 5 wherein the selected bac-
25 terial strain is a strain of Leuconostoc oenos.
7. A method according to claim 6 wherein the strain of Leuconostoc oenos is a strain selected from the group consisting of DSM 7008, DSM 7009, DSM 7010, DSM 7011, DSM 7012, DSM 7013, DSM 7014 and DSM 7015.

8. A method according to claim 1 wherein the selected bacterial strain has at least one of following characteristics:

(a) capable of decarboxylating malic acid in wine at a pH of 3.2 or lower,

5 (b) capable of decarboxylating malic acid in wine in the presence of 25 mg SO₂ per L or more,

(c) capable of decarboxylating malic acid in wine having an ethanol content of 10 vol% or higher,

10 (d) a survival rate of at least 80% when introduced into a wine having a pH of 3.2 or lower and containing at least 10 vol% ethanol,

15 (e) a survival rate of at least 50% when introduced into a wine having a pH of 3.2 or lower and containing at least 25 mg SO₂ per L and at least 12 vol% ethanol,

(f) resistant to attack by bacteriophages, or

(g) capable of retaining at least one of the characteristics (a) through (f) during propagation and concentration.

20 9. A method according to claim 8 wherein the selected bacterial strain has at least two of the characteristics (a) through (g).

10. A method according to claim 9 wherein the selected bacterial strain has at least three of the characteristics (a) through (g).

25

11. A method according to claim 10 wherein the selected bacterial strain has all of the characteristics (a) through (g).
12. A method according to claim 1 wherein the provided concentrate has a content of colony forming units which is in the range of 10^9 to 10^{12} per g.
13. A method according to claim 12 wherein the concentrate has a content of colony forming units which is in the range of 10^{11} to 10^{12} per g.
14. A method according to claim 1 wherein the concentrate is preserved.
- 15 A method according to claim 14 wherein the preserved concentrate is frozen.
16. A method according to claim 14 wherein the preserved concentrate is freeze-dried.
17. A method according to claim 1 wherein the concentrate is added in an amount resulting in the introduction into the wine or fruit juice of a number of the malolactically active bacteria which is in the range of 10^5 to 10^7 CFU per mL.
18. A method according to claim 17 wherein the number of the malolactically active bacteria being introduced into the wine or the fruit juice is in the range of 10^6 to 10^7 .
19. A method according to claim 18 wherein the number of the malolactically active bacteria being introduced into the wine or the fruit juice is in the range of 5×10^6 to 10^7 .
20. A method according to claim 1 wherein the wine or fruit juice has a content of malic acid which is at least 4 g per L.

21. A method according to claim 20 wherein the period of time to obtain the resulting malolactically fermented wine or fruit juice is at the most 15 days.
22. A method according to claim 21 wherein the period of time
5 to obtain the resulting malolactically fermented wine or fruit juice is at the most 10 days.
23. A method according to claim 22 wherein the period of time to obtain the resulting malolactically fermented wine or fruit juice is at the most 8 days.
- 10 24. A method according to claim 1 wherein the wine or fruit juice has a content of malic acid which is at least 5.5 g per L.
25. A method according to claim 24 wherein the period of time to obtain the resulting malolactically fermented wine or
15 fruit juice is at the most 15 days.
26. A method according to claim 25 wherein the period of time to obtain the resulting malolactically fermented wine or fruit juice is at the most 10 days.
27. A method according to claim 26 wherein the period of time
20 to obtain the resulting malolactically fermented wine or fruit juice is at the most 8 days.
28. A method according to claim 1 wherein the obtained malolactically fermented wine or fruit juice has a content of malic acid which is at the most 0.1 g per L.
- 25 29. A selected strain of a malolactically active bacterium which; when it is added to a wine or fruit juice in a frozen or freeze-dried state at a concentration of CFUs which is in the range of 1×10^6 to 5×10^7 per mL of the wine or fruit juice, is capable of reducing at least 4 g of malic acid per
30 L of wine or fruit juice to less than 0.5 g per L within a

period of time which is at the most 15 days, said strain having at least one of which is a strain having at least one of following characteristics:

- 5 (a) capable of decarboxylating malic acid in wine at a pH of 3.2 or lower,
- (b) capable of decarboxylating malic acid in wine in the presence of 25 mg SO₂ per L or more,
- (c) capable of decarboxylating malic acid in wine having an ethanol content of 10 vol% or higher,
- 10 (d) a survival rate of at least 80% when introduced into a wine having a pH of 3.2 or lower and containing at least 10 vol% ethanol,
- (e) a survival rate of at least 50% when introduced into a wine having a pH of 3.2 or lower and contain-
15 ing at least 25 mg SO₂ per L and at least 12 vol% ethanol,
- (f) resistant to attack by bacteriophages, or
- (g) capable of retaining at least one of the characteristics (a) through (f) during propagation and
20 concentration.

30. A strain according to claim 29 which is a strain having at least two of the characteristics (a) through (g).

31. A strain according to claim 30 which is a strain having at least three of the characteristics (a) through (g).

25 32. A strain according to claim 31 which is a strain having all of the characteristics (a) through (g).

33. A strain according to claim 29 which is one selected from the group consisting of a Leuconostoc species, a Lactobacillus species and a Pediococcus species.

34. A strain according to claim 33 which is a strain of
5 Leuconostoc oenos.

35. A strain according to claim 34 wherein the strain of Leuconostoc oenos is selected from the group consisting of DSM 7008, DSM 7009, DSM 7010, DSM 7011, DSM 7012, DSM 7013, DSM 7014 and DSM 7015.

10 36. A concentrate of a selected strain of a malolactically active bacterium as defined in claim 29.

37. A concentrate according to claim 36 which is a preserved concentrate.

15 38. A concentrate according to claim 37 which is a frozen concentrate.

39. A concentrate according to claim 37 which is a freeze-dried concentrate.

20 40. A concentrate according to claim 36 which is a concentrate having a content of colony forming units being in range of 10^9 to 10^{12} per g.

41. A concentrate according to claim 40 which is a concentrate having a content of colony forming units being in the range of 10^{11} to 10^{12} per g.

25 42. A concentrate according to claim 36 which is a concentrate, the direct introduction of which into a wine containing at least 4 g of malic acid per L, at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 15 days.

43. A concentrate according to claim 42 which is a concentrate, the direct introduction of which into a wine containing at least 4 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 10 days.

44. A concentrate according to claim 43 which is a concentrate, the direct introduction of which into a wine containing at least 4 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 8 days.

45. A concentrate according to claim 36 which is a concentrate, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 15 days.

46. A concentrate according to claim 45 which is a concentrate, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 10 days.

47. A concentrate according to claim 46 which is a concentrate, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L in less than 8 days.

48. A malolactic starter culture composition comprising a concentrate of a selected malolactically active bacterial strain, as defined in claim 36 and at least one further

ingredient selected from cryoprotectants, bacterial nutrients and bulking agents.

49. A composition according to claim 48 which is preserved.

50. A composition according to claim 49 which is a frozen
5 composition.

51. A composition according to claim 49 which is a freeze-dried composition.

52. A composition according to claim 48 which is a composition having a content of colony forming units being in range
10 of 10^9 to 10^{12} per g.

53. A composition according to claim 52 which is a composition having a content of colony forming units being in the range of 10^{11} to 10^{12} per g.

54. A composition according to claim 48 which is a composition,
15 the direct introduction of which into a wine containing at least 4 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time
20 which is at the most 15 days.

55. A composition according to claim 54 which is a composition, the direct introduction of which into a wine containing at least 4 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the
25 wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time which is at the most 10 days.

56. A composition according to claim 55 which is a composition, the direct introduction of which into a wine containing
30 at least 4 g of malic acid per L at a concentration of CFUs

which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time which is at the most 8 days.

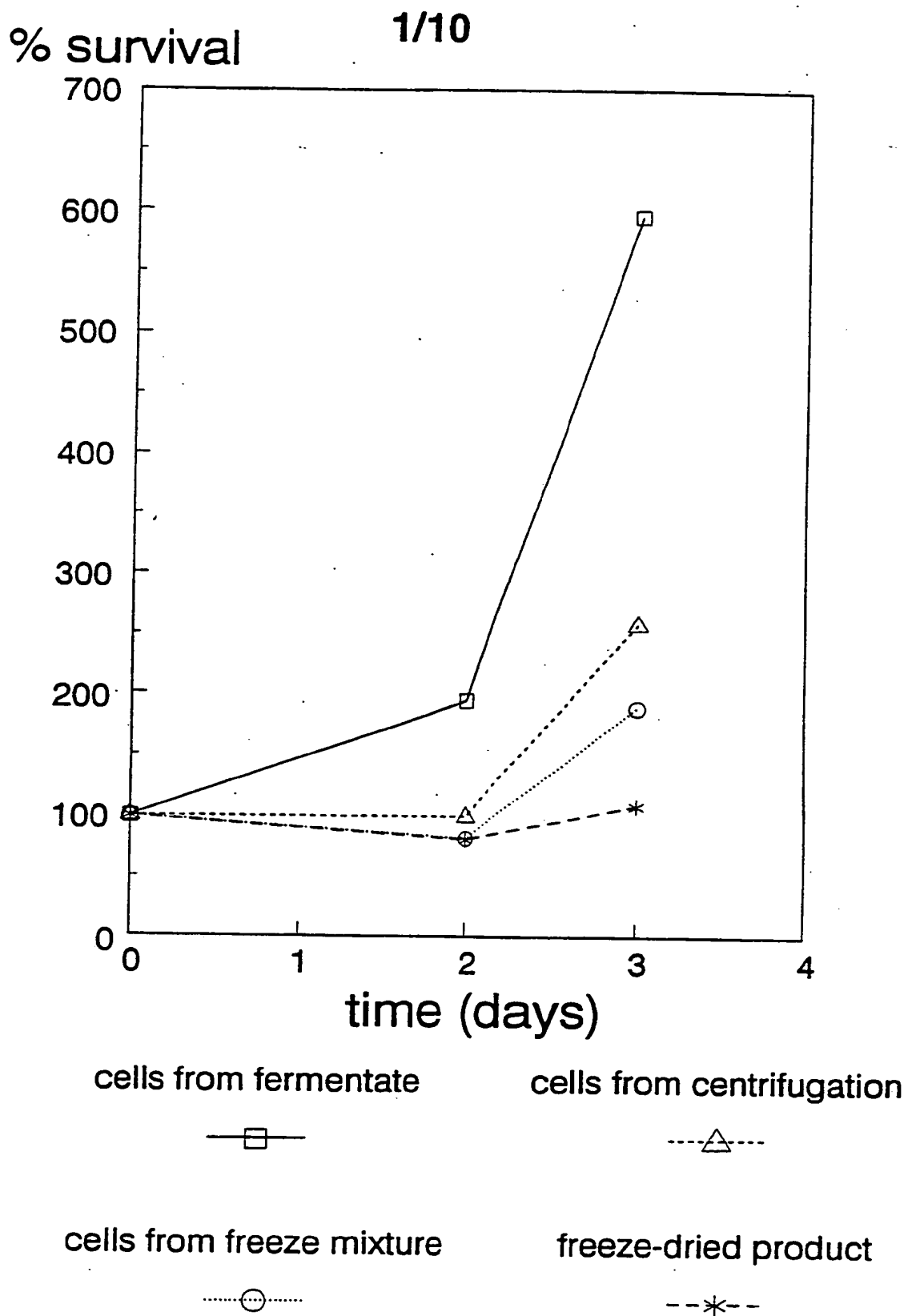
- 5 57. A composition according to claim 48 which is a composition, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing
10 at the most 0.5 g malic acid per L within a period of time which is at the most 15 days.

58. A composition according to claim 57 which is a composition, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs
15 which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time which is at the most 10 days.

59. A composition according to claim 58 which is a composition, the direct introduction of which into a wine containing at least 5.5 g of malic acid per L at a concentration of CFUs which is in the range of 5×10^6 to 5×10^7 per mL of the wine, results in a malolactically fermented wine containing at the most 0.5 g malic acid per L within a period of time
20 which is at the most 8 days.

60. A composition according to claim 48 comprising a multiplicity of selected malolactically active bacteria.

61. A composition according to claim 48 wherein the cryoprotectant is gelatine.

**Fig. 1**

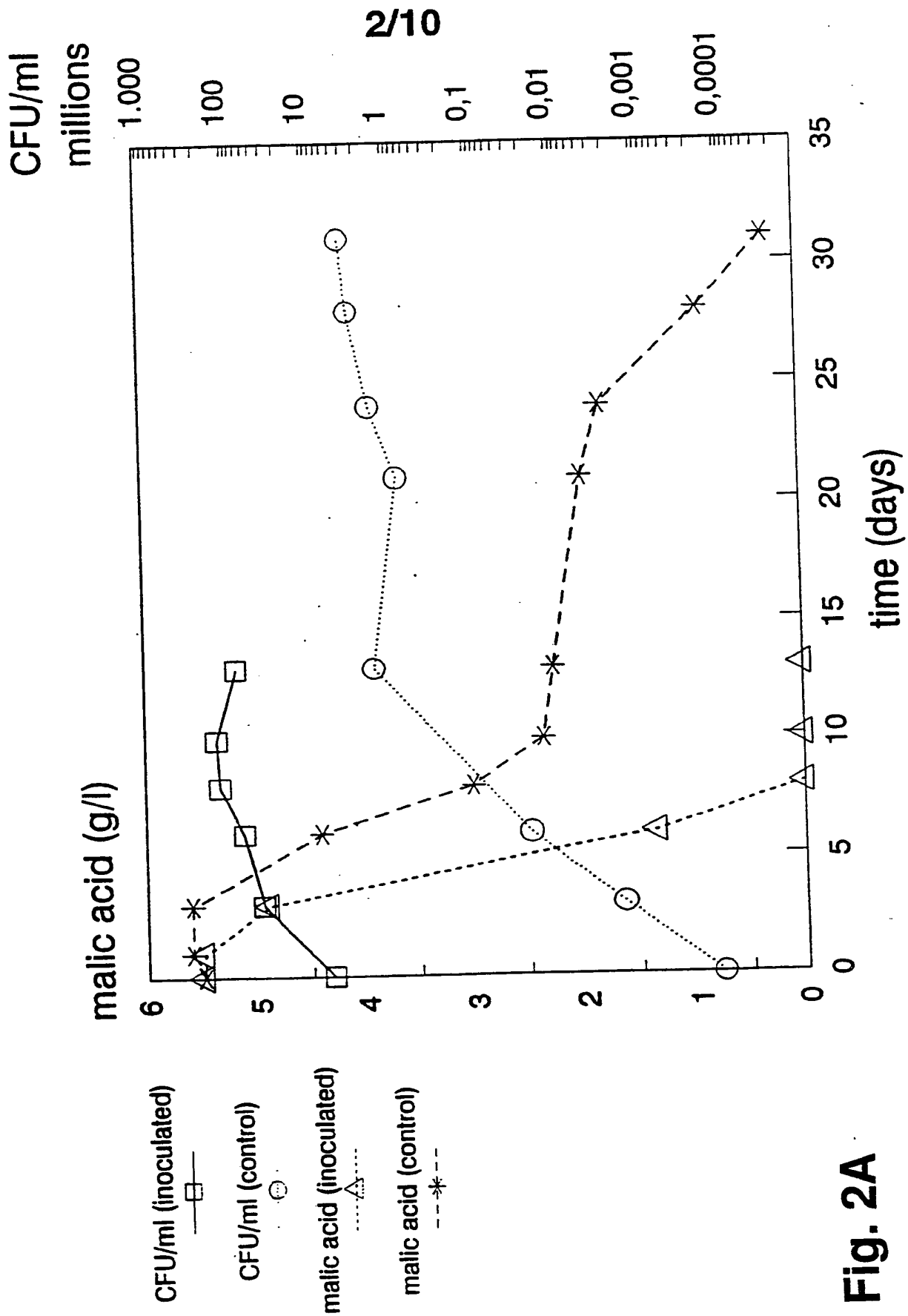
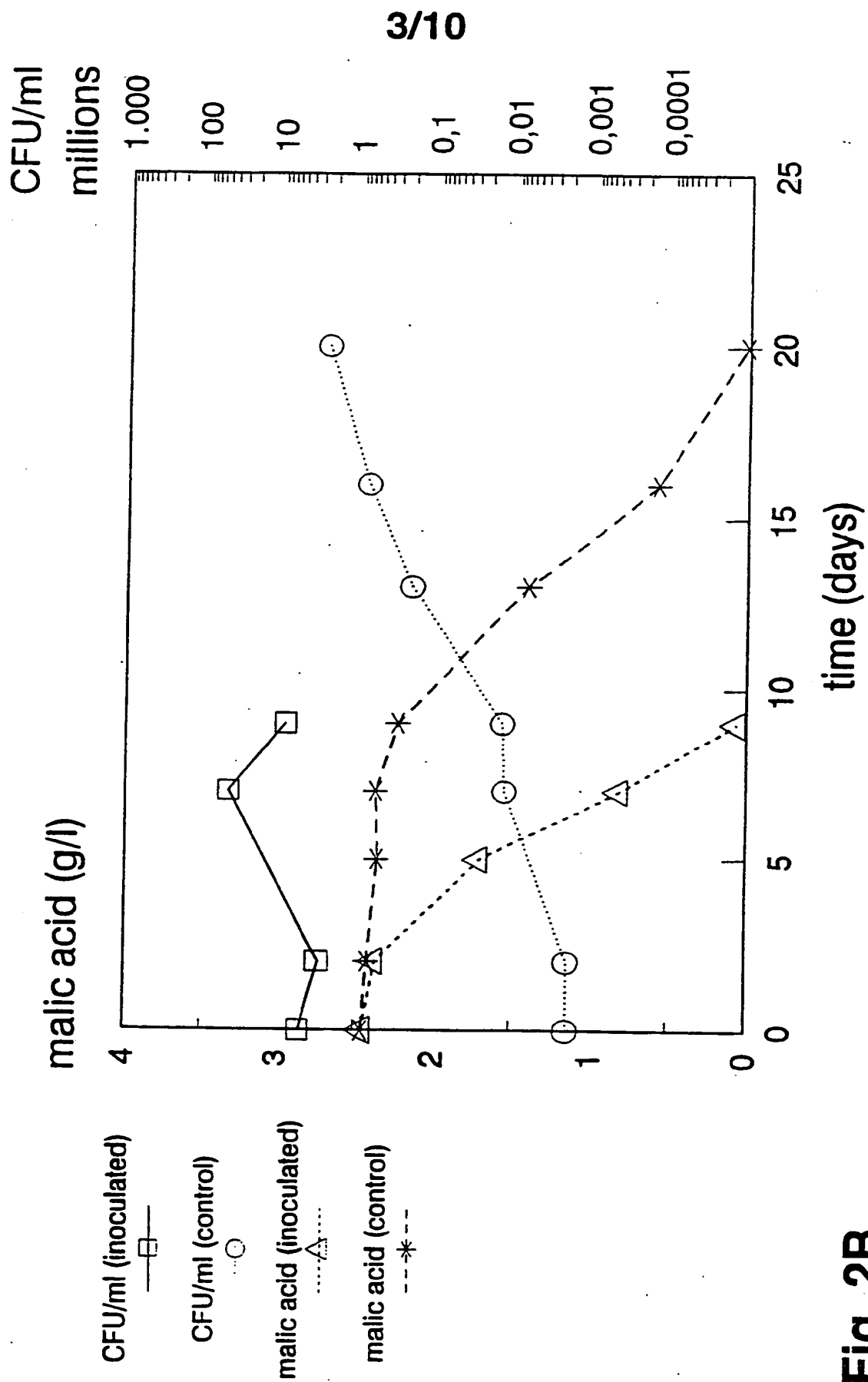
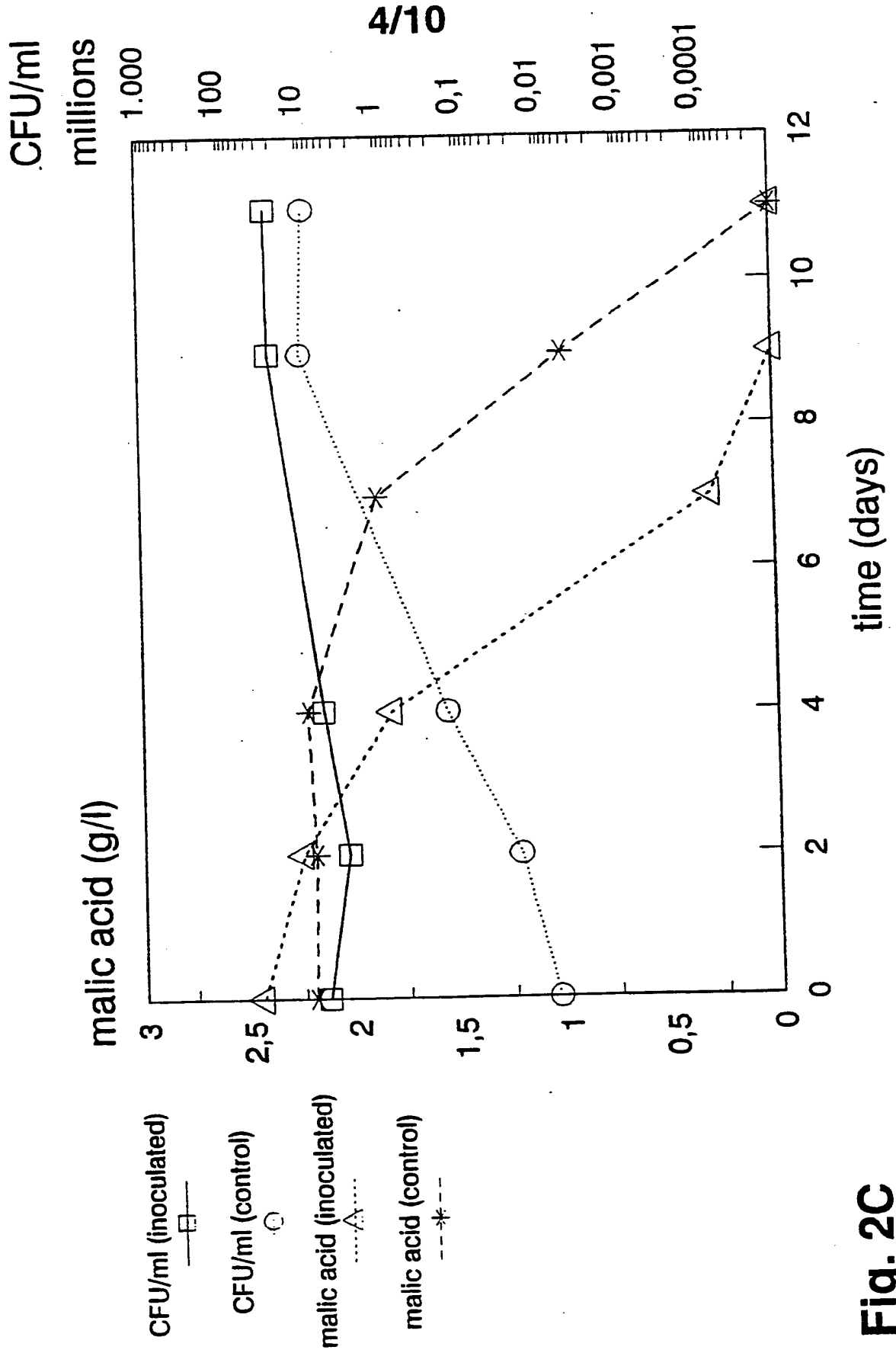


Fig. 2A

**Fig. 2B**

**Fig. 2C**

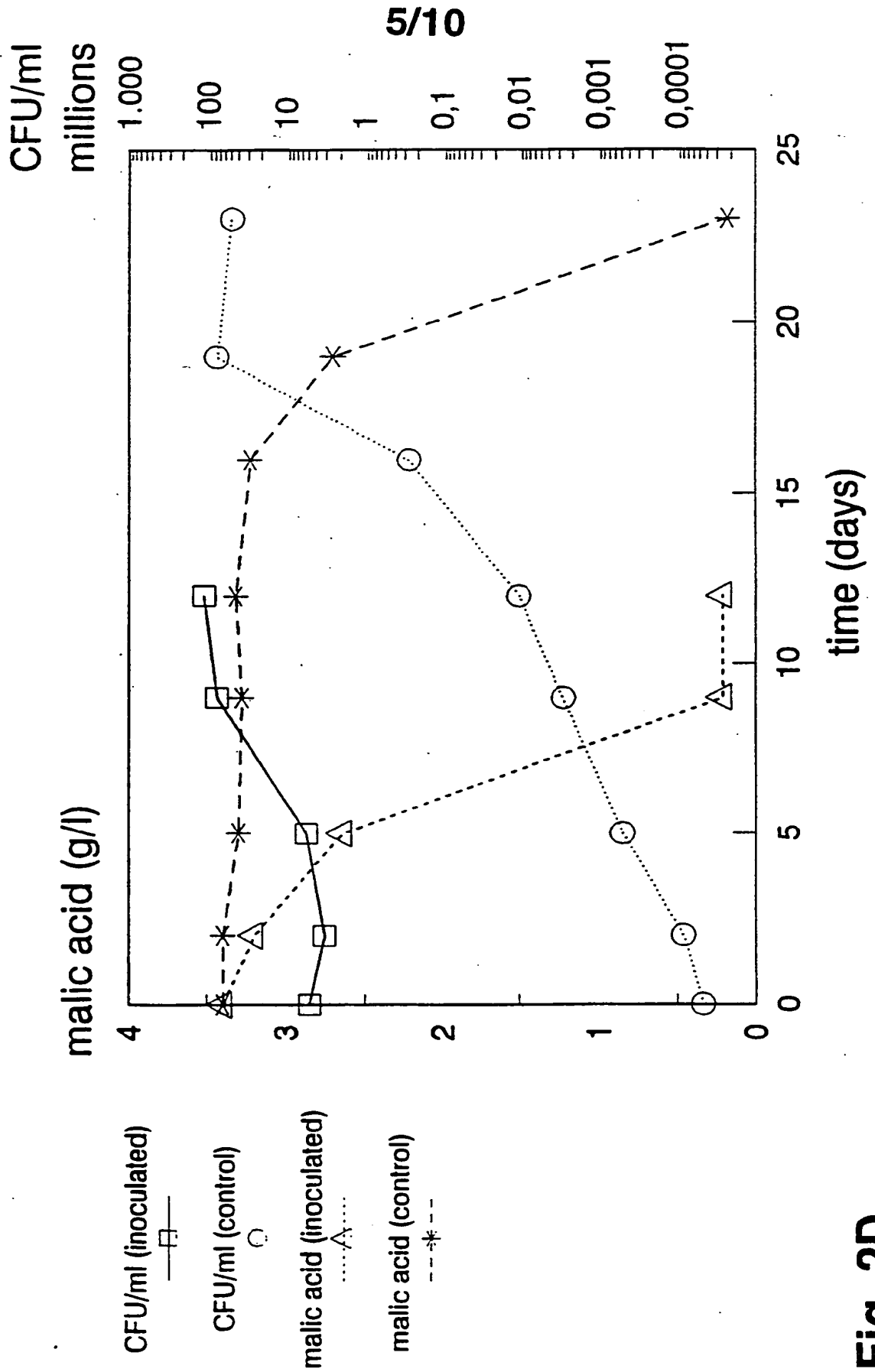


Fig. 2D

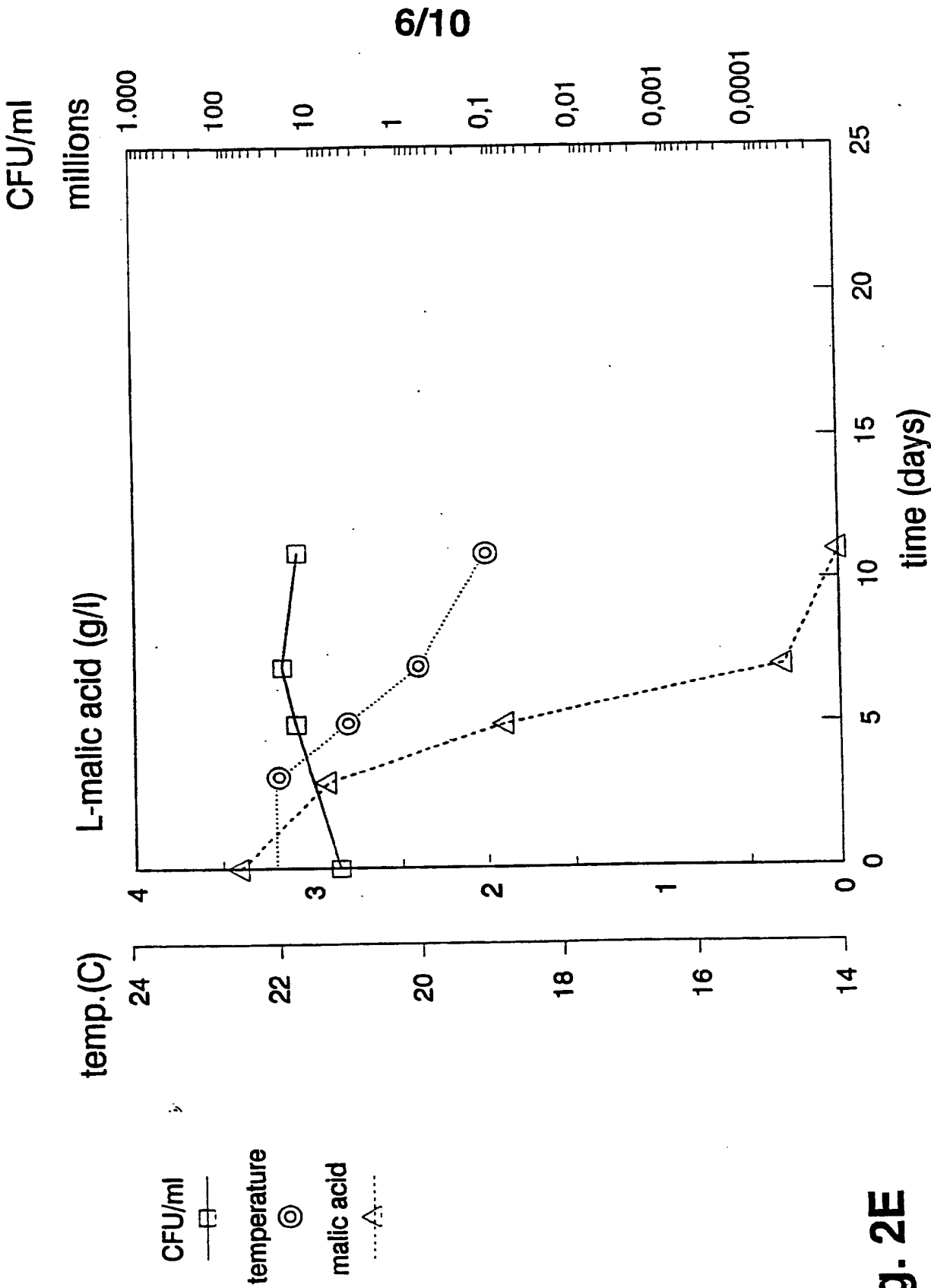


Fig. 2E

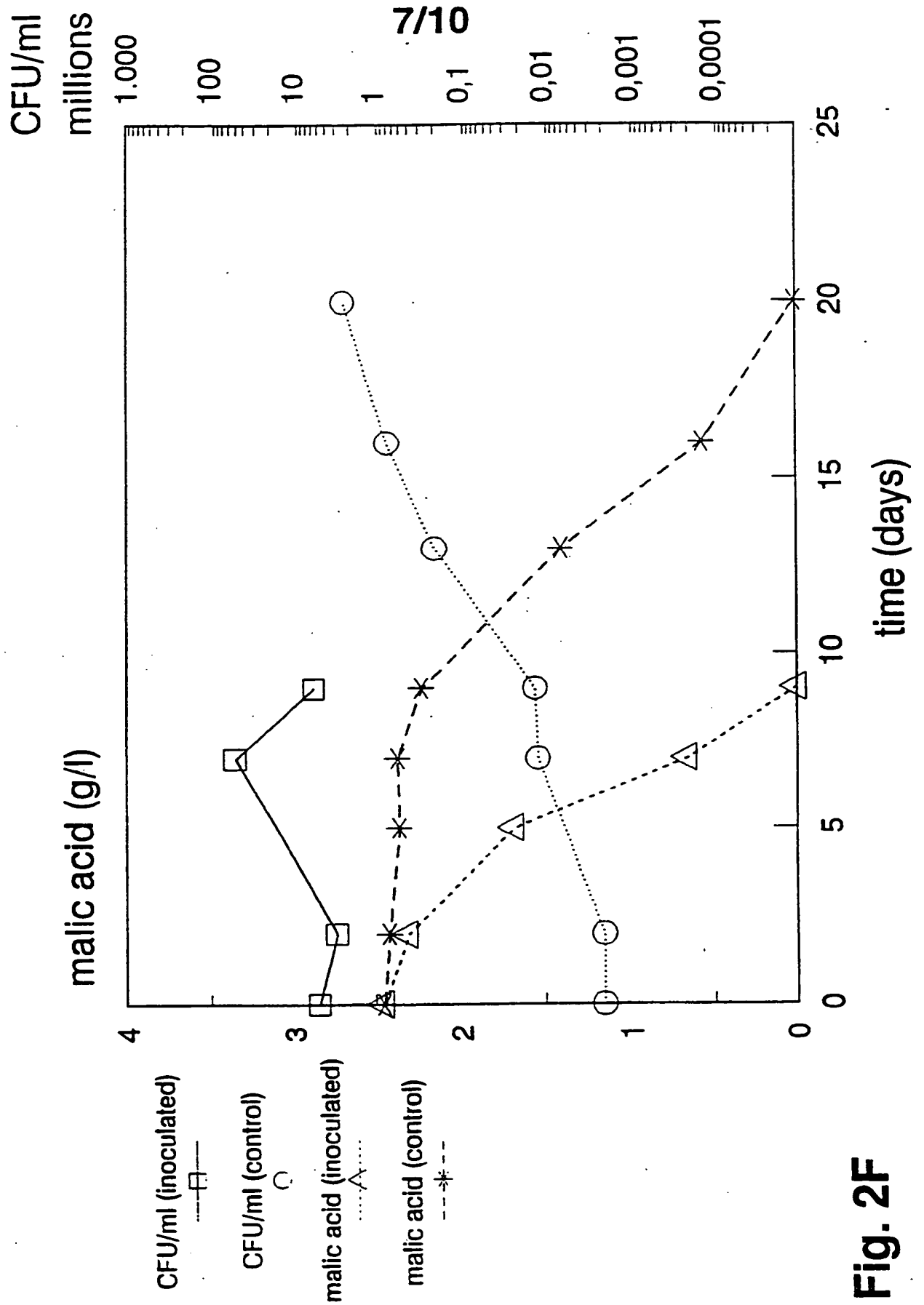


Fig. 2F

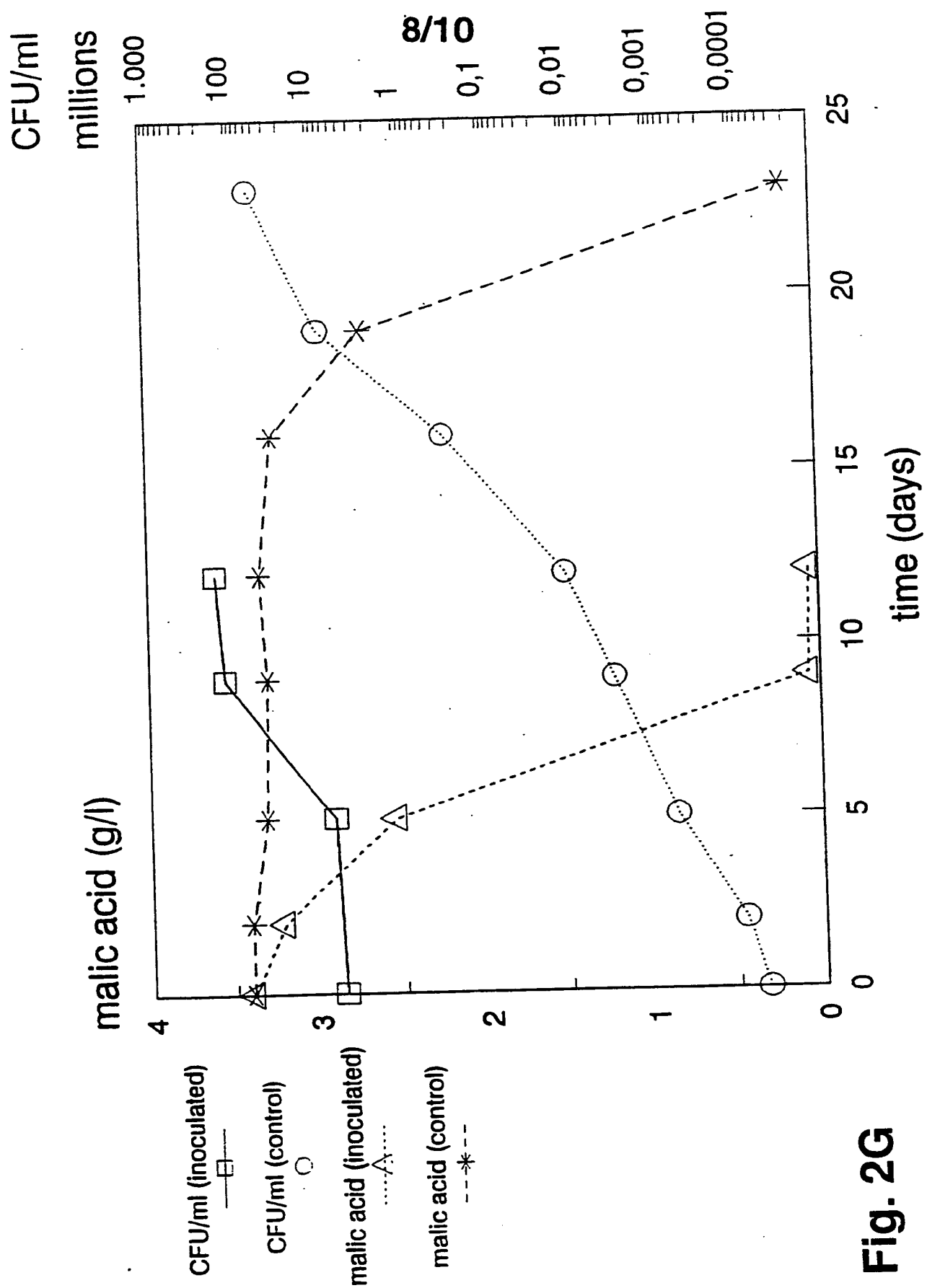


Fig. 2G

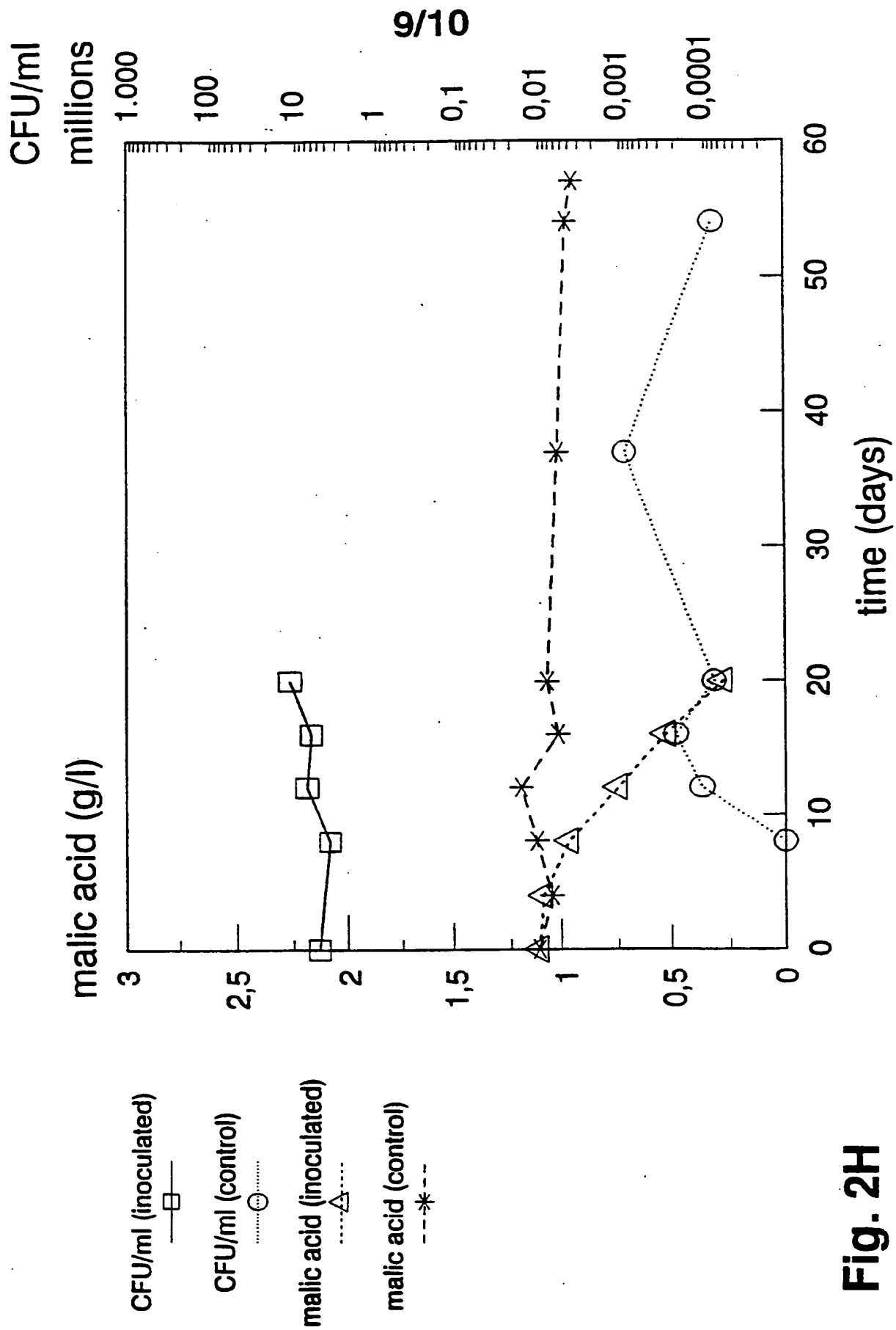


Fig. 2H

10/10

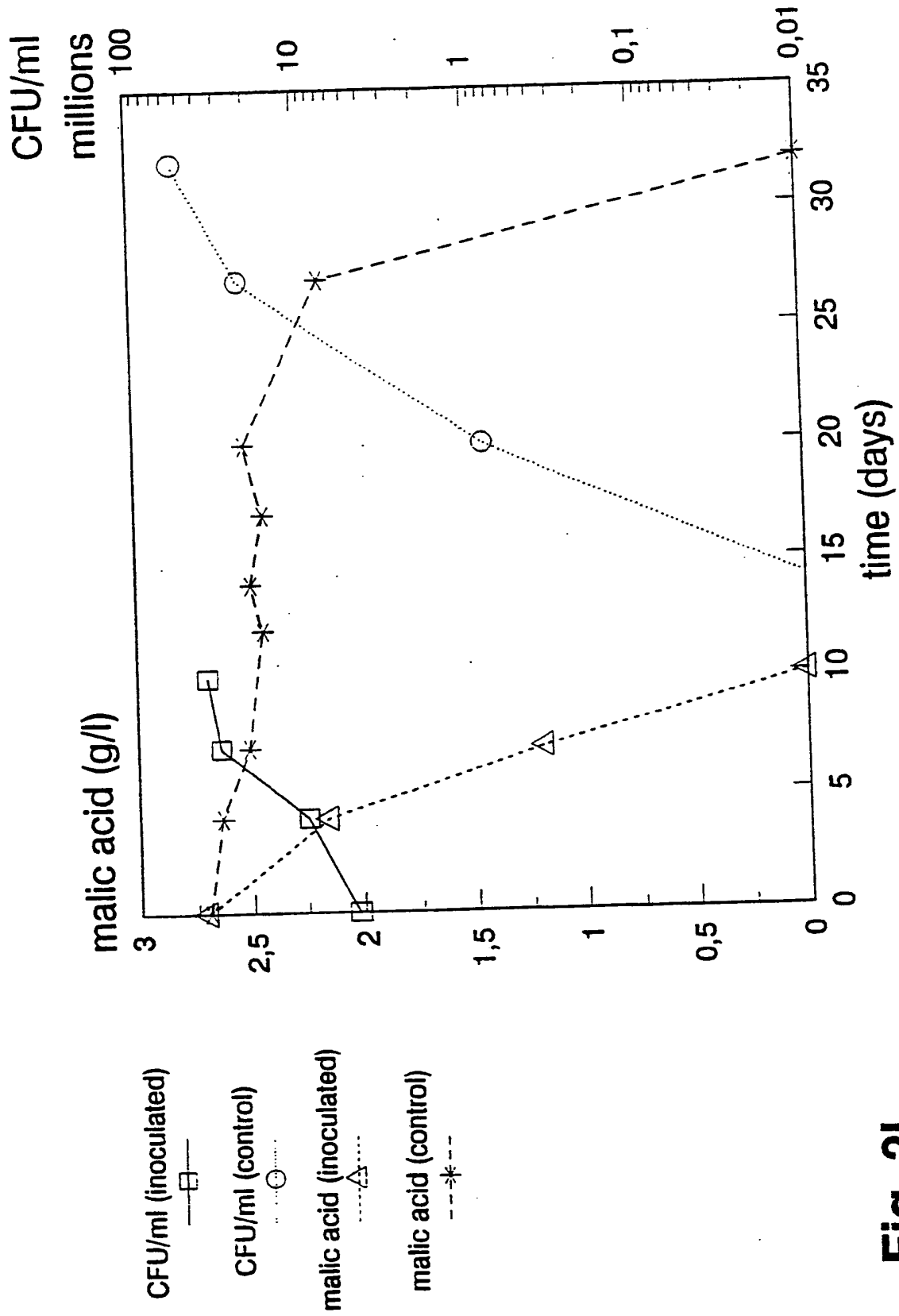


Fig. 2I

INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 93/00116

A. CLASSIFICATION OF SUBJECT MATTER

IPC5: C12G 1/02, C12H 1/00, C12P 7/56

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC5: C12G, C12H, C12P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPI, WPIL, CLAIMS, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FR, A, 2485037 (SILVER JACK ET MCNICHOLL FRANCES), 24 December 1981 (24.12.81), page 2, line 36 - line 37; page 4, line 14; page 13, example 3; page 15, example 8; claim 14	1-9,11-30, 32-61
Y	--	10,31
X	WO, A1, 8906685 (CHR. HANSEN'S LABORATORIUM A/S), 27 July 1989 (27.07.89), the claims	1-5,8,12-29, 33,36-61
Y	--	9-10,30-31

☒ Further documents are listed in the continuation of Box C.☒ See patent family annex.

* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

1 July 1993

Date of mailing of the international search report

08-07-1993

Name and mailing address of the ISA/
Swedish Patent Office
Box 5055, S-102 42 STOCKHOLM
Facsimile No. +46 8 666 02 86

Authorized officer

Carolina Palmcrantz
Telephone No. +46 8 782 25 00

INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 93/00116

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Dialog Information Services, File 5, Biosis, Dialog accession no. 5324127, Biosis number 81091434, Davis C R et al: "Growth and meta- bolism of lactic-acid bacteria during and after malolactic fermentation of wines at different pH", Appl Environ Microbiol 51 (3), 1986, 539-545 --	9-10,30-31
Y	Dialog Information Services, File 5, Biosis, Dialog accession no. 5235886, Biosis number 81003193, Davis C et al: " Occurrence and properties of bacteriophages of leucono- stoc-oenos in australian wines", Appl Environ Microbiol 50 (4), 1985, 872-876 --	9-10,30-31
Y	Dialog Information Services, File 5, Biosis, Dialog accession no. 7430956, Biosis number 89081975, Britz T J et al: "The combination effect of pH sulfur dioxide ethanol and temperature on the growth of leuconostoc-oenos", J Appl Bacteriol 68 (1), 1990, 23-32 --	9-10,30-31
A	EP, A1, 141878 (MICROLIFE TECHNICS, INC.), 22 May 1985 (22.05.85), page 3, line 11 - line 13; page 4, table I; claims 1,3 --	1-61
A	Dialog Information Services, File 351, WPI, Dialog accession no. 008250107, WPI accession no. 90-137108/18, SNOW BRAND MILK PRODUCTS: "New lactobacilli SBT 0568 belonging to genus leuconostoc - used to accelerate malo-lactic fermentation to improve wine flavour", JP 2086765, A, 900327, 9018 (Basic) -----	1-61

INTERNATIONAL SEARCH REPORT
Information on patent family members

28/05/93

International application No.

PCT/DK 93/00116

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
FR-A- 2485037	24/12/81	AU-A- 7204981 CH-A,B- 662819 DE-A- 3124404	07/01/82 30/10/87 11/03/82
WO-A1- 8906685	27/07/89	AU-A- 3054489 EP-A- 0398957	11/08/89 28/11/90
EP-A1- 141878	22/05/85	NONE	

Form PCT/ISA/210 (patent family annex) (July 1992)

THIS PAGE BLANK (USPTO)